

# DNA Barcode Library for Selected Species of Korean Tenebrionidae (Coleoptera)

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## Abstract

Members of the family Tenebrionidae (Coleoptera) show varying degrees of morphological differences within a species, as well as between species. In this study, we selected 15 species (in 9 genera, 7 tribes, and 4 subfamilies) of Korean Tenebrionidae belonging to 4 functional groups depending on the feeding habits and habitats, and constructed their DNA barcode library composed of 658-bp partial mitochondrial cytochrome c oxidase subunit I (COI) genes. In addition, we tested the efficiency of DNA barcodes for identifying the intraspecific and interspecific differences within members of the family. Our results indicated that the COI genes are effective in identification of the Tenebrionidae species.

**Key words:** Tenebrionidae, DNA barcode library, COI gene, habitat preference, Korea

## Introduction

The family Tenebrionidae or darkling beetles is one of the most diverse groups in Coleoptera. Approximately 22,000 species belonging to this family have been described worldwide, and 129 species and 66 genera are known in Korea (Jung 2011). Members of the family show extensive variations with respect to body size (1–50 mm), form, and vestiture. Most adults of the Tenebrionidae have elongated oval bodies; heads with horns sometimes, antennae in different shapes such as filiform, moniliform, serrate, and rarely flagellate, mesocoxae with exposed trochantins, tarsal formula are usually 5-5-4, tarsal claws are not pectinate, and intersegmental membranes of abdomen are exposed between ventrites 3–5 (Jung 2011). The larvae of Tenebrionidae are called false wireworms because of the presence of elongated, glabrous, and strongly sclerotized body.

Both adults and larvae of Tenebrionidae inhabit diverse habitats or microhabitats such as grain products, rotten wood, fungi, soil, and sand dunes (Arnett *et al.* 2002, Bouchard *et al.* 2005, Löbl *et al.* 2008, Jung 2011). They show adaptive inhabitation in the roots of plants, under the barks, in rotten wood, in mushrooms, and under leaf debris (Jung 2008). Watt (1974) classified Tenebrionidae into the following 4 groups depending on the feeding habits and habitats:

(A) Cereal group: This group belongs to the tribes Tenebrionini, Triboliini, and genus *Alphitobius* and are pests of stored cereal products. The most important species under

this group are *Tribolium confusum* Duval and *Tribolium castaneum* Herbst, which cause considerable damage to stored cereals, especially in temperate climate zones (Cotton 1956, Watt 1974, Grimaldi & Engel 2005).

(B) Forest group associated with rotten wood: Members are the species *Cerogria janthinipennis* (Fairmaire) from the subfamily Lagriinae which live under barks of logs, and some other Lagriini species which live on ground, flowers, and grass (Watt 1974, Hayashi 1964). The species from the tribe Amarygmini are also associated with dead and rotten wood. They are found under the bark and associated subcortical space of logs and are usually nocturnal.

(C) Macrofungi group: Most beetles belonging to the subfamilies of Diaperinae (genera *Platydema*, *Basanus*, and *Scaphidema*) and Stenochiinae (Tribe Cnodalonini) specialize in feeding on macrofungi such as Basidiomycetes, which is the most common food for fungivorous tenebrionids in rotten wood (Marshall 1982, Leschen 1990, Kim & Jung 2005, Jung & Kim 2008, Jung & Lee 2011, Jung & Lee 2012). Both larvae and adults of fungivorous tenebrionids spend their whole lives in the fruiting bodies of bracket fungi.

(D) Sand dune group: Many tenebrionids have developed specific adaptations that allow them to inhabit sand dunes in deserts, riversides, and coastal dunes (Krivolutskaya 1973, Watt 1974, Grimaldi & Engel 2005, Jung *et al.* 2009). In particular, members of the tribe Opatrini (genus *Gonocephalum*) adapted to hot and dry sand (Kim & Kim 2000). Members of this group show a high degree of interspecific

variations as well as a substantial level of intraspecific similarities in morphology.

Identification of insect species through traditional taxonomy primarily relies on external morphology. However, phenotypic variation in species with taxonomically similar traits can pose considerable difficulties in species identification (Ball & Armstrong 2006). DNA barcode is a useful tool to solve taxonomic issues, particularly if a comprehensive DNA barcode library is available. Species identification is considered successful when the test DNA sequence matches most closely with its congeners in the library data. However, a DNA barcode reference library is not available for the Korean Tenebrionidae, and misidentified sequences are sometimes existent in the present GenBank.

In this study, we selected 15 species of Korean Tenebrionidae according to feeding habits and habitats shown above, and constructed a DNA barcode library using specific regions of the COI genes. The efficiency of this library was tested for identifying of the Korean Tenebrionidae species.

## Materials and methods

### Specimens

Specimens used in this study were collected from throughout South Korea during 2008–2012. The specimens were collected from diverse habitats such as rotten trees covered with lichen and sand dunes (Table 1) and were examined and photographed using a microscope (Carl Zeiss Discovery V12) with an image analyzer program (AxioCam I Cc1, Germany).

Adult specimens preserved in 95% ethanol were used for the mitochondrial DNA analysis. We used 50 specimens for DNA extraction, and voucher specimens were deposited in the Entomological Museum of Korea University (KU), Seoul.

The abbreviations used in this study are as follows: SL (Seoul), GG (Gyeonggi-do), CB (Chungcheongbuk-do), CN (Chungcheongnam-do), GB (Gyeongsangbuk-do), GN (Gyeongsangnam-do), GW (Gangweon-do), JN (Jeollanam-do), and JJ (Jeju-do).

### DNA extraction, PCR amplification and DNA sequencing

DNA was extracted from the thorax muscle or legs of adult specimen by using the DNeasy Blood and Tissue Kit (QIAGEN Genomics Inc., Dusseldorf, Germany), according to the manufacturer's protocol. PCR amplification was performed under the following conditions: initial denaturation at 94°C for 3 min; 40 cycles at 94°C for 30 sec (denaturation), 48°C for 1 min (annealing), 72°C for 30 sec (extension), and a final extension at 72°C for 10 min. The PCR products were maintained at 4°C. We used 2 µL of template DNA in a total reaction volume of 20 µL of PCR reaction mixture that con-

tained ultrapure water, 0.5 U Taq DNA polymerase in 1 × Taq buffer containing 20 mM Tris-HCl (pH 8.2), 50 mM KCl, 4 mM MgCl<sub>2</sub>, 200 µM total dNTPs, and 0.5 µM forward and reverse primers. For PCR, we used the universal primer pair LCO1490 (Forward) (5'-GGTCAACAAATCA TAAAGATATTGG-3') and HCO2198 (Reverse) (5'-TAAA CTCAGGGTGACCAAAAAATCA-3') to amplify the 5'-end of the COI gene (Folmer *et al.* 1994). The PCR products were visualized on 1.5% agarose gel stained with GelRed™ (Biotium Inc, Hayward, CA, USA). The PCR products were then sequenced using an ABI 3730 Automated Sequencer (Macrogen Co., Seoul, Korea). The sequences were assembled and aligned using the software, BioEdit (version 7.0.5.3, Hall 1999, <http://www.mbio.ncsu.edu/BioEdit/bioedit.html>).

### Data analysis

Mitochondrial COI sequences were aligned using Clustal X (Version 2.0, Larkin 2007) and edited to a final length of > 600 bp. Pairwise nucleotide sequence divergences, which was calculated using the Kimura two-parameter (K2P) model (Kimura 1980), and neighbor-joining (NJ) analysis, which was performed using the software Mega 5 (Tamura *et al.* 2011), were used to examine relationships among the taxa. All sequences obtained in this study were deposited in the GenBank.

## Results

### Taxonomic accounts

Taxonomic accounts dealt in this study are based on previous studies (Jung 2005, 2008, 2011, Jung *et al.* 2009) with modifications and additions of figures and collecting localities. Synonyms are based on previous catalogs (Jung 2011); species authorities and their literatures are therefore not included in the reference.

Family Tenebrionidae

#### 1. *Cerogria janthinipennis* (Fairmaire) (Fig. 1A)

*Lagria janthinipennis* Fairmaire, 1886: 349.

*Cerogria antennata* Jacobson, 1913: 1016; ZSK, 1968: 112; ESK & KSAE, 1994: 176; Kwon *et al.*, 1996: 162.

*Cerogria janthinipennis*: Jacobson, 1913: 1016; Kim *et al.*, 1994: 101; ESK & KSAE, 1994: 176; Kim, 1995: 129; Kim, 1996: 8; Kim & Kim, 1996: 43; Kim *et al.*, 1999: 125; Kim, 2000: 127; Kim, 2002: 225; Kim *et al.*, 2002: 115.

*Cerogria* notability: Park *et al.*, 1993: 153 (Misidentification of *Cerogria janthinipennis*).

**Table 1.** Details of Tenebrionidae specimens used for DNA barcoding and analysis in this study

Species name	Collection locality	Habitat	GenBank accession number (COI)
<i>Cerogria janthinipennis</i> (Fairmaire)	GB	Leaf litter, Rotten wood	KC440128
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407728
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407729
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407730
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407731
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407732
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407733
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407734
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407735
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407736
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407737
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407738
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407739
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407740
<i>Tenebrio molitor</i> Linnaeus	Seoul	Cereal	KC407741
<i>Tribolium castaneum</i> (Herbst)	NCBI	Cereal	NC003081
<i>Tribolium castaneum</i> (Herbst)	NCBI	Cereal	FM877932
<i>Tribolium castaneum</i> (Herbst)	GN	Cereal	KC440129
<i>Tribolium confusum</i> Jacquelin Du Val.	GN	Cereal	JQ350711
<i>Tribolium confusum</i> Jacquelin Du Val.	GN	Cereal	KC407742
<i>Tribolium confusum</i> Jacquelin Du Val.	GN	Cereal	KC407743
<i>Plesiophthalmus davidis</i> Fairmaire	GW	Dead and rotten wood	KC440130
<i>Plesiophthalmus davidis</i> Fairmaire	GW	Dead and rotten wood	KC440131
<i>Gonocephalum persimile</i> (Lewis)	GW	Soil, Sand dunes	KC440132
<i>Gonocephalum pubens</i> (Marseul)	CN	Soil, Sand dunes	KC440133
<i>Gonocephalum pubens</i> (Marseul)	CN	Soil, Sand dunes	KC440134
<i>Gonocephalum pubens</i> (Marseul)	GW	Soil, Sand dunes	KC440135
<i>Gonocephalum pubens</i> (Marseul)	GW	Soil, Sand dunes	KC440136
<i>Gonocephalum coenosum</i> Kaszab	GN	Soil, Sand dunes	KC440137
<i>Gonocephalum coenosum</i> Kaszab	GN	Soil, Sand dunes	KC440138
<i>Gonocephalum coriaceum</i> (Motschulsky)	GB	Soil, Sand dunes	KC440139
<i>Basanus tsushimensis</i> M.T. Chûjô	Seoul	Macrofungi in rotten wood	KC440140
<i>Basanus tsushimensis</i> M.T. Chûjô	Seoul	Macrofungi in rotten wood	KC440141
<i>Basanus tsushimensis</i> M.T. Chûjô	Seoul	Macrofungi in rotten wood	KC440142
<i>Platydema recticorne</i> Lewis	GG	Macrofungi in rotten wood	KC440143
<i>Platydema recticorne</i> Lewis	GG	Macrofungi in rotten wood	KC440144
<i>Platydema recticorne</i> Lewis	Seoul	Macrofungi in rotten wood	KC440145
<i>Platydema recticorne</i> Lewis	Seoul	Macrofungi in rotten wood	KC440146
<i>Platydema fumosum</i> Lewis	GG	Macrofungi in rotten wood	KC440147
<i>Platydema fumosum</i> Lewis	GG	Macrofungi in rotten wood	KC440148
<i>Platydema fumosum</i> Lewis	Known	Macrofungi in rotten wood	KC440149
<i>Platydema nigroaeneum</i> Motschulsky	GG	Macrofungi in rotten wood	KC440150
<i>Platydema nigroaeneum</i> Motschulsky	GG	Macrofungi in rotten wood	KC440151
<i>Platydema nigroaeneum</i> Motschulsky	Seoul	Macrofungi in rotten wood	KC440152
<i>Platydema nigroaeneum</i> Motschulsky	Seoul	Macrofungi in rotten wood	KC440153
<i>Diaperis lewisi lewisi</i> Bates	GG	Macrofungi in rotten wood	KC440154
<i>Promethis valgipes valgipes</i> (Marseul)	GG	Macrofungi in rotten wood	KC440155
<i>Promethis valgipes valgipes</i> (Marseul)	GG	Macrofungi in rotten wood	KC440156
<i>Promethis valgipes valgipes</i> (Marseul)	JJ	Macrofungi in rotten wood	KC440157
<i>Promethis valgipes valgipes</i> (Marseul)	JJ	Macrofungi in rotten wood	KC440158

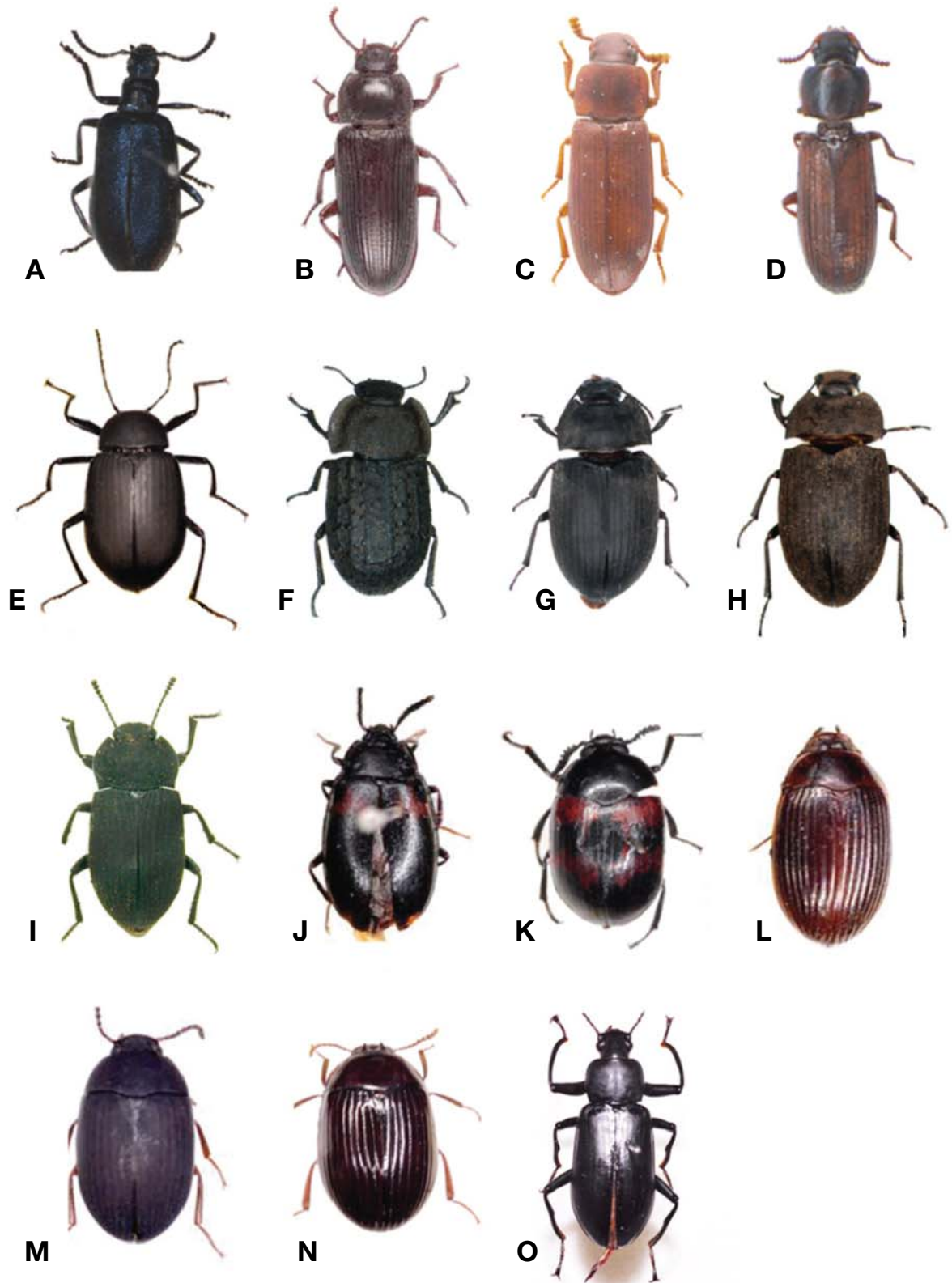
**Specimen examined.** 1 adult. GB: Uljin-gun, Jukbyeon-myeon, Bongpyeong-ri, 28.V.2012, HJ Park [KU].

**Distribution.** Korea, Japan, China, East Siberia.

**Habitat preference.** Under bark of logs, on the ground on flowers and grass (Watt 1974).

## 2. *Tenebrio molitor* Linnaeus (Fig. 1B)

*Tenebrio molitor* Linnaeus, 1758: 417; Kim *et al.*, 1994: 178; Kim & Oh, 1990: 130; Kim, 1995: 407; Kwon & Choi, 1986: 109–110; Kwon *et al.*, 1996: 163; Löbl *et al.*, 2008: 299; Yoon *et al.*, 1989: 112.



**Figure 1.** Habitus of Tenebrionidae. A. *Cerogria janthinipennis*; B. *Tenebrio molitor*; C. *Tribolium castaneum*; D. *Tribolium confusum*; E. *Plesio-phthalmuns davidis*; F. *Gonocephalum persimile*; G. *Gonocephalum pubens*; H. *Gonocephalum coenosum*; I. *Gonocephalum coriaceum*; J. *Basanus tsushimensis*; K. *Diaperis lewisi lewisi*; L. *Platydema recticorne*; M. *Platydema fumosum*; N. *Platydema nigroaeneum*; O. *Promethis valgipes valgipes*.

**Specimen examined.** 14 adults. SL: Seongbuk-gu, Anam-dong, 17.IV.2012, SY Cho [KU].

**Distribution.** Cosmopolitan.

**Habitat preference.** Stored cereal products.

### 3. *Tribolium castaneum* (Herbst) (Fig. 1C)

*Colydium castaneum* Herbst, 1797: 282.

*Tribolium ferrugineum* Fabricius: Cho *et al.*, 1969: 309; Gu, 1970: 66.

*Tribolium castaneum* (Herbst): Chûjô & Lee, 1992: 39; Kim *et al.*, 1994: 177; Kim, 1995: 407; Kwon *et al.*, 1996: 163; Masumoto, 1984: 13.

**Specimen examined.** 1 adult. GN: Yangsan-si, Ugok-dong, 19.III.2012, JH Na [KU].

**Distribution.** Cosmopolitan.

**Habitat preference.** Stored cereal products.

### 4. *Tribolium confusum* Jacquelin Du Val (Fig. 1D)

*Tribolium confusum* Jacquelin Du Val, 1868: 181; Kim *et al.*, 1994: 177; Kwon *et al.*, 1996: 163; Löbl *et al.*, 2008: 301.

**Specimen examined.** 3 adults. GN: Yangsan-si, Eogok-dong, 19.III.2012, JH Na [KU].

**Distribution.** Cosmopolitan.

**Habitat preference.** Stored cereal products.

### 5. *Plesiophthalmus davidis* Fairmaire (Fig. 1E)

*Plesiophthalmus davidis* Fairmaire, 1868: 121; Chûjô & Lee, 1993: 119; Kim *et al.*, 1994: 178; Kwon *et al.*, 1996: 163.

**Specimen examined.** 2 adults. GW: Pyeongchang-gun, Jinbu-myeon, Mt. Odaesan, 18.V.2012, WY Choi [KU].

**Distribution.** Korea, China.

**Habitat preference.** Dead and rotten wood or under the bark of dead trees.

### 6. *Gonocephalum persimile* (Lewis) (Fig. 1F)

*Opatrum persimile* Lewis, 1894: 381.

*Gonocephalum persimile*: Chûjô, 1963: 152–153; Chûjô & Baba, 1979: 50; Chûjô & Lee, 1993: 109–110; ESK & KSAE, 1994: 176; Gebien, 1910b: 324, 1939: 447; Hua Li, 2002: 139; Iwan & Löbl, 2008: 265; Kaszab, 1952a: 624; 1952b: 680; 1968: 8; Kim, 1981a: 65; 1995: 141; Kwon & Choi, 1986: 106; Kwon *et al.*, 1996: 163; Masumoto, 1985: 35; Reichardt, 1936: 113; Schuster, 1928: 990.

**Specimen examined.** 1 adult. GW: Youngwol-gun, Kimsatgat-myeon, Yemil-ri, 5.V.2012, AY Kim [KU].

**Distribution.** Palaearctic: Korea, Japan, Mongolia, NE China, SE Siberia.

**Habitat preference.** Sand dune and soil under stones.

### 7. *Gonocephalum pubens* (Marseul) (Fig. 1G)

*Opatrum pubens* Marseul, 1876: 97.

*Opatrum pubens* Mars. Var. *obtusicolle*: Cho, 1957: 52; Heyden, 1887: 259; Kolbe, 1886: 95; Okamoto, 1924: 183.

*Gonocephalum pubens*: Champion, 1895: 115; Cho, 1963: 50; Chûjô, 1966: 8; Chûjô et Baba, 1979: 50; Chûjô & Lee, 1992: 33–34; 1993: 110; Chûjô, 1994: 152–153; ESK & KSAE, 1994: 176; Gebien, 1910b: 324, 1913: 4, 1939: 447; Gu Gun, 1973: 34–67; Hua Li, 2002: 139; Iwan & Löbl, 2008: 265; Kaszab, 1941a: 51, 1952a: 594, 1952b: 679; Kim & Kim, 1972: 157; Kim, 1995: 406; Kim & Lee, 1979: 83; Kim, 1980: 142, 1981a: 65, 1983: 83, 1995: 141; Kwon & Choi, 1986: 106; Kwon *et al.*, 1996: 163; Lee *et al.*, 1985: 408; Masumoto, 1985: 35–36; Reichardt, 1936: 111; Schuster, 1928: 990.

**Specimen examined.** 4 adults. CN: Taean-gun, Changfi-ri, Gijipo beach, 5.V.2012, SY Cho & SH Han [KU]; KW: Kangreung-si, Anhyun-dong, Sungeut beach. 3–4.VIII.2010. HG Min & GS Oh [KU].

**Distribution.** Palaearctic: Korea, China, India, Japan, Taiwan.

**Habitat preference.** Sand dune.

### 8. *Gonocephalum coenosum* Kaszab (Fig. 1H)

*Gonocephalum coenosum* Kaszab, 1952: 643–646; Chûjô, 1963: 152–153; Chûjô, 1966: 6, 1978b: 1; Chûjô & Imsaka, 1982: 59; Chûjô & Lee, 1992: 32; ESK & KSAE, 1994: 176; Hua Li, 2002: 138; Iwan & Löbl, 2008: 263; Kim, 1983: 83; Kim, 1995: 405; Kim & Oh, 1991: 155; Kwon & Choi, 1986: 106; Kwon *et al.*, 1996: 163; Lee & Kwon, 1974: 43; Lee *et al.*, 1985: 408; Masumoto, 1985: 35–36; Masumoto & Kondo, 1984: 3; Matsumoto & Akita, 2008: 121.

**Specimen examined.** 2 adults. JN: Shinan-gun, Bigeum-myeon, Duksan-ri, Bigeum-do, 28.VII.2011, BH Jung & JB Seung [KU].

**Distribution.** Palaearctic: Korea, China, Japan, Taiwan.

**Habitat preference.** Sand dune.

### 9. *Gonocephalum coriaceum* (Motschulsky) (Fig. 1I)

*Opatrum coriaceum* Motschulsky, 1858: 34; Cho, 1957: 52; Kolbe, 1886: 201; Lewis, 1894: 381.

*Gonocephalum coriaceum*: Chûjô, 1963: 152–153; Chûjô, 1994: 247; Chûjô & Baba, 1979: 49–50; Chûjô & Lee, 1992: 33; 1994: 189; ESK & KSAE, 1994: 176; Gebien,

1910b: 322, 1913: 6, 1939: 447; Harold, 1876: 130; Hua Li, 2002: 138; Iwan & Löbl, 2008: 263; Kaszab, 1941a: 51, 1952a: 667, 1952b: 451, 1954b: 248; Kim *et al.*, 1972: 222; Kim & Lee, 1979: 83; Kim, 1981b: 344; 1983: 83; Kim, 1995: 405; Kwon & Choi, 1986: 106; Kwon *et al.*, 1996: 163; Marseul, 1876: 96; Masumoto, 1985: 35–36; Masumoto & Kondo, 1984: 4; Motschulsky, 1866: 173; Reichardt, 1936: 112; Reitter, 1904: 143; Schuster, 1928: 989; Yoon & Nam, 1978: 83.

**Specimen examined.** 1 adult. GB: Ulgin-gun, Seo-myeon, Hawon-ri, Bulyoungsa, 02.VIII.2006, BH Jung [KU].

**Distribution.** Korea, China, Japan, NE.China, Taiwan.

**Habitat preference.** Sand dune and soil under stones.

### 10. *Basanus tsushimensis* Chûjô (Fig. 1J)

*Basanus tsushimensis* M.T. Chûjô, 1963: 17; Kwon & Choi, 1986: 105; Chûjô & Lee, 1992: 31; Chûjô & Lee, 1993: 109; Paik *et al.*, 1995: 405; Kwon *et al.*, 1996: 162; Kim *et al.*, 2004: 111; Kim & Jung, 2005: 95.

*Basanus tsushimensis kompancevi* Kaszab & Medvedev, 1984: 75; ESK & KSAE, 1994: 176.

**Specimen examined.** 3 adults. SL: Gangbuk-gu, Ui-dong, Dosunsa, 22.VI.2006, AY Kim [KU].

**Distribution.** Korea, Japan.

**Habitat preference.** Fruiting bodes of bracket fungi.

### 11. *Diaperis lewisi lewisi* Bates (Fig. 1K)

*Diaperis lewisi lewisi* Bates, 1873: 14; Kolbe, 1886: 202; Kim *et al.*, 1972: 103; Kwon & Choi, 1986: 108; Chûjô & Lee, 1993: 109; ESK & KSAE, 1994: 176; Kwon *et al.*, 1996: 163; Kim & Jung, 2005: 95.

**Specimen examined.** 1 adult. SL: Gangbuk-gu, Ui-dong, Dosunsa, 22.VI.2006, AY Kim [KU].

**Distribution.** Korea, Japan, China (Manchuria), Russia (Southern Primorye, Siberia).

**Habitat preference.** Fruiting bodes of bracket fungi.

### 12. *Platydemia recticorne* Lewis (Fig. 1L)

*Platydemia recticorne* Lewis, 1894: 394; Chûjô & Lee, 1993: 109; Chûjô & Lee, 1994: 189; Kwon *et al.*, 1996: 162; Kim & Jung, 2005: 9.

**Specimen examined.** 4 adults. GG: Namyangju-si, Jinjeop-myeon, Bupyeong-ri, Gwangreung, 18.IX.2009, BH Jung [KU]; SL: Gangdong-gu, Gil-dong, Gildong Eco-park, 22.V.2010, BH Jung [KU].

**Distribution.** Korea, Japan.

**Habitat preference.** Fruiting bodes of bracket fungi with rotten tree.

### 13. *Platydemia fumosum* Lewis (Fig. 1M)

*Platydemia fumosum* Lewis, 1894: 395; Chûjô & Lee, 1993: 109; Kwon *et al.*, 1996: 162; Kim & Jung, 2005: 9.

**Specimen examined.** 3 adults. GG: Guri-si, Donggu-dong, Donggureung, 30.VII.2012, BH Jung [KU].

**Distribution.** Korea, Japan (Honshu, Tsushima Island).

**Habitat preference.** Fruiting bodes of bracket fungi with rotten tree.

### 14. *Platydemia nigroaeneum* Motschulsky (Fig. 1N)

*Platydemia nigroaeneum* Motschulsky, 1860: 18; Heyden, 1887: 243; Kim *et al.*, 1971: 157; Kim & Nam, 1981: 126; Kwon & Choi, 1986: 105; ESK & KSAE, 1994: 176; Kim *et al.*, 1994: 127; Kwon *et al.*, 1996: 162; Kim *et al.*, 1998: 163; Kim *et al.*, 2002: 115; Kim & Jung, 2005: 9; Kim *et al.*, 2005: 223.

**Specimen examined.** 4 adults. GG: Gwangju-si, Chowol-eup, Mt. Mugapsan, 27.V.2010, BH Jung [KU]; Guri-si, Donggu-dong, Donggureung, 30.VII.2012, BH Jung [KU]; SL: Gangdong-gu, Gil-dong, Gildong Eco-park, 22.V.2010, BH Jung [KU].

**Distribution.** Korea, Japan.

**Habitat preference.** Fruiting bodes of bracket fungi.

### 15. *Promethis valgipes valgipes* (Marseul) (Fig. 1O)

*Nyctobates valgipes* Marseul, 1876: 117.

*Setenis valgipes*: Lewis, 1894: 473; Kwon & Choi, 1986: 106.

**Table 2.** Summary of intraspecific Kimura 2-parameter distances in the Korean Tenebrionidae species

Scientific name	Mean	Range	S.E.	No. of specimens
Subfamily Lagriinae				
<i>Cerogria janthinipennis</i>	n/c	n/c	n/c	1
Subfamily Tenebrioninae				
<i>Tenebrio molitor</i>	1.17	0.00–2.39	0.38	14
<i>Tribolium castaneum</i>	0.72	0.72–0.72	0.34	3
<i>Tribolium confusum</i>	0.00	0.00–0.00	0.00	3
<i>Plesiophthalmus davidis</i>	0.00	0.00–0.00	0.00	2
<i>Gonocephalum persimile</i>	n/c	n/c	n/c	1
<i>Gonocephalum pubens</i>	0.18	0.00–0.36	0.15	5
<i>Gonocephalum coenosum</i>	0.00	0.00–0.00	0.00	2
<i>Gonocephalum coriaceum</i>	n/c	n/c	n/c	1
Subfamily Diaperinae				
<i>Basanus tsushimensis</i>	0.00	0.00–0.00	0.00	3
<i>Platydemia recticorne</i>	0.18	0.00–0.36	0.16	4
<i>Platydemia fumosum</i>	0.36	0.36–0.36	0.25	3
<i>Platydemia nigroaeneum</i>	0.36	0.00–0.72	0.22	4
<i>Diaperis lewisi lewisi</i>	n/c	n/c	n/c	1
Subfamily Stenochiinae				
<i>Promethis valgipes valgipes</i>	0.09	0.00–0.18	0.08	4

**Table 3.** Pairwise comparison of COI gene sequences in Tenebrionidae. The values were calculated using the Kimura 2-parameter method (Kimura, 1980). All results are based on the gene divergence analysis of 15 Korean species of Tenebrionidae according to habitat preference\*

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
1. <i>Cerogria janthinipennis</i>															
2. <i>Tenebrio molitor</i>	0.299														
3. <i>Tribolium castaneum</i>	0.300	0.222													
4. <i>Tribolium confusum</i>	0.249	0.231	0.208												
5. <i>Plesiophthalmus davidis</i>	0.291	0.226	0.200	0.190											
6. <i>Gonocephalum persimile</i>	0.257	0.224	0.203	0.186	0.193										
7. <i>Gonocephalum pubens</i>	0.239	0.238	0.167	0.208	0.204	0.154									
8. <i>Gonocephalum coenosum</i>	0.239	0.238	0.167	0.208	0.204	0.154	0.001								
9. <i>Gonocephalum coriaceum</i>	0.226	0.219	0.187	0.188	0.213	0.150	0.122	0.122							
10. <i>Basanus tsushimensis</i>	0.302	0.290	0.271	0.262	0.238	0.259	0.225	0.225	0.246						
11. <i>Platydemus recticorne</i>	0.274	0.210	0.201	0.218	0.201	0.186	0.167	0.167	0.171	0.255					
12. <i>Platydemus fumosum</i>	0.239	0.219	0.180	0.212	0.229	0.214	0.176	0.176	0.190	0.265	0.171				
13. <i>Platydemus nigroaeneum</i>	0.247	0.219	0.185	0.180	0.194	0.158	0.167	0.167	0.175	0.234	0.155	0.168			
14. <i>Diaperis lewisi lewisi</i>	0.286	0.242	0.207	0.218	0.211	0.197	0.190	0.190	0.204	0.228	0.211	0.207	0.195		
15. <i>Promethis valgipes valgipes</i>	0.270	0.207	0.203	0.225	0.221	0.210	0.215	0.216	0.212	0.263	0.217	0.217	0.243	0.226	

\*Habitat preference: in cereal (genera *Tenebrio* and *Tribolium* in tribe Tenebrionini); in sand dune (genus *Gonocephalum* in tribe Opatrini); in macrofungi (genera *Basanus* and *Platydemus* in tribe Diaperinae, *Promethis valgipes valgipes* in tribe Stenochiinae); in rotten wood (*Cerogria janthinipennis* in tribe Lagriinae and *Plesiophthalmus* in tribe Amarygmni)

**Table 4.** Base composition and GC percentage of COI gene sequences in the Korean Tenebrionidae specie determined in this study

Scientific name	Sequence length	%A	%C	%G	%T	%G+C	%A+T	No. of specimens
<i>Cerogria janthinipennis</i>	658.0	41.3	14.7	28.0	16.0	42.7	57.3	1
<i>Tenebrio molitor</i>	658.0	28.9	22.2	30.9	18.0	53.1	46.9	14
<i>Tribolium castaneum</i>	652.2	31.6	21.9	29.9	16.6	51.8	48.2	3
<i>Tribolium confusum</i> Jacquelin.	658.0	33.0	19.6	30.9	16.6	50.5	49.5	3
<i>Plesiophthalmus davidis</i>	658.0	30.4	21.6	32.5	15.5	54.1	45.9	2
<i>Gonocephalum persimile</i>	658.0	33.6	19.8	29.8	16.9	49.5	50.5	1
<i>Gonocephalum pubens</i>	658.0	34.5	19.4	29.8	16.3	49.2	50.8	4
<i>Gonocephalum coenosum</i>	653.0	34.5	19.5	29.8	16.3	49.3	50.7	2
<i>Gonocephalum coriaceum</i>	658.0	35.9	18.1	30.1	16.0	48.2	51.8	1
<i>Basanus tsushimensis</i>	658.0	32.1	23.6	29.3	15.0	52.9	47.1	3
<i>Platydemus recticorne</i>	620.9	31.1	22.0	30.6	16.3	52.6	47.4	4
<i>Platydemus fumosum</i>	652.7	32.9	20.9	29.9	16.3	50.8	49.2	3
<i>Platydemus nigroaeneum</i>	636.5	33.0	20.2	30.6	16.1	50.8	49.2	4
<i>Diaperis lewisi lewisi</i>	658.0	33.0	22.6	29.2	15.2	51.8	48.2	1
<i>Promethis valgipes valgipes</i>	658.0	29.6	23.3	29.4	17.7	52.6	47.4	4
Average	652.1	31.6	21.4	30.3	16.8	51.7	48.3	50

*Promethis valgipes*: Kaszab, 1988: 85; Chûjô & Lee, 1992: 31; ESK & KSAE, 1994: 176; Kwon *et al.*, 1996: 162.

**Specimen examined.** 4 adults. JJ: Seogwipo-si, Seongsan-eup, 5.V.2012, BH Jung & JB Seung [KU]; Jeji-si, Gujwa-eup, Pyeongdae-ri, Bijarim, 18.VIII.2012, BH Jung & JB Seung [KU].

**Distribution.** Korea, Japan, China.

**Habitat preference.** Fruiting bodies of bracket fungi.

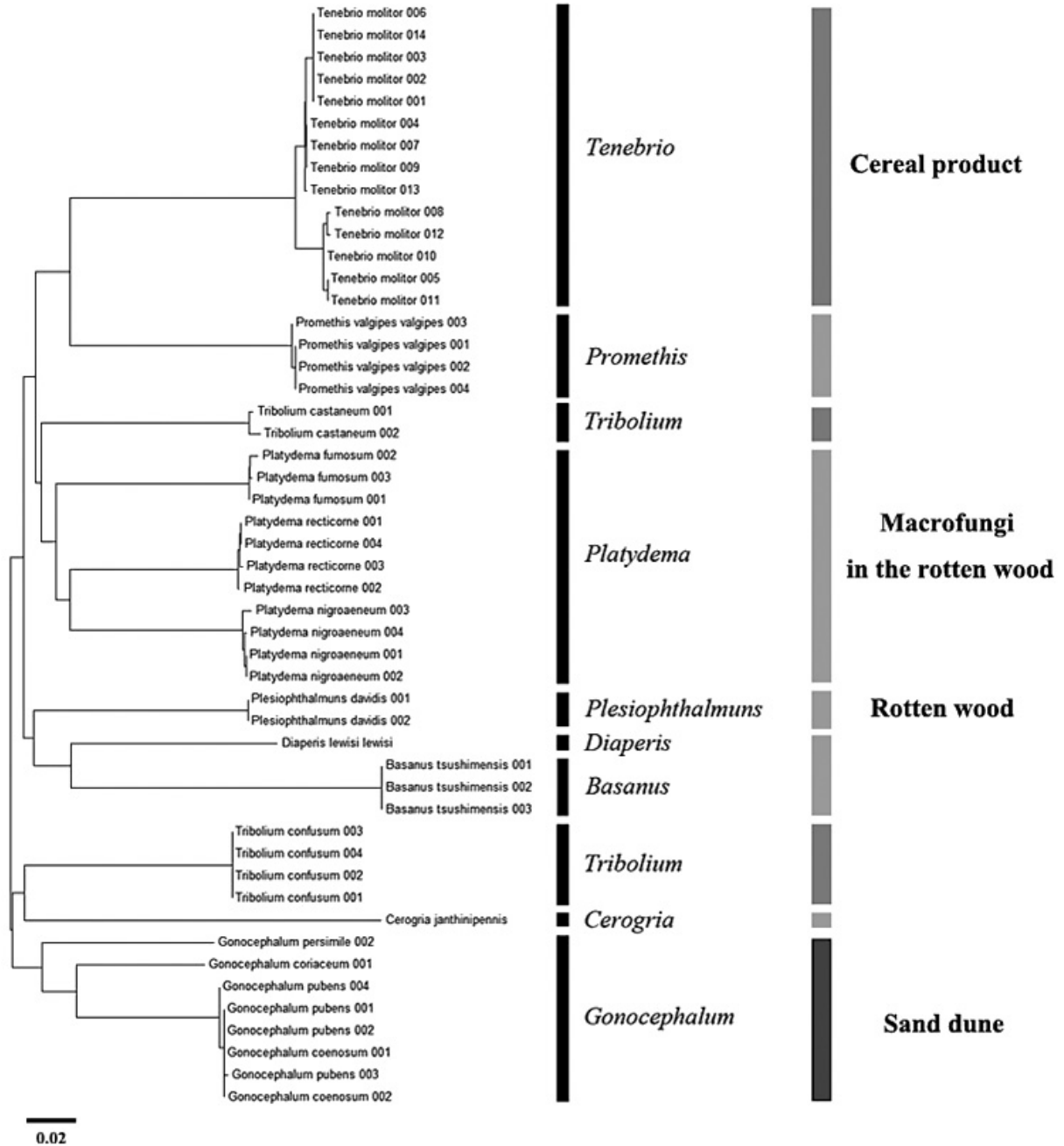
### Construction of DNA barcode library

The 50 COI sequences ranged from 600 to 658 bp (mean=652 bp with <1% ambiguity) in length from 50 specimens

belonging to 15 species, 9 genera, and 4 subfamilies.

The pairwise genetic distances among the individuals within one species are shown in Table 2. The mean of intra-specific divergence variation was 0.9% and ranged between 0.0–2.39%. The result matched the range for the species *Tenebrio molitor*.

The mean of interspecific genetic divergence was 19.5% (SE=1.3%), whereas divergence variation widely ranged from a minimum of 0.1% to a maximum of 30.2% (Table 3). Majority of the species (65%) exceeded a value of 20% genetic divergence between the species. When the sequences of 2 species, *Cerogria janthinipennis* (Lagriinae) and *Basanus tsushimensis* (Diaperinae), were compared for genetic divergence, their DNA barcode gaps were far larger (mean=



**Figure 2.** Phylogenetic trees using neighbor-joining analyses for COI divergence in the selected Korean Tenebrionidae species.

30.2%, SE=2.7%) as compared to that between other species (mean=19.5%, SE=1.3%).

The nucleotide content of the COI gene sequences determined in this study, covering the Tenebrionidae, is summarized in Table 4. The COI sequences from the sampled species showed a strong A+T bias in the nucleotide content (mean=58.3%) relative to the C+G content (mean=41.7%), which is typical of arthropods. Only *Cerogria janthinipennis*

*nis* showed a low G+C content of 42.7%. The G+C content of the genus *Gonocephalum*, which inhabits sand dunes, was 49.5% (range 48.2–49.5%). The mean G+C content of genus *Platydema* was 51.4% (range 50.8–52.6%), while the G+C content of 2 other species *Basanus tsushimensis* and *Diaperis lewisi lewisi*, which occupy similar habitats were 52.9% and 51.8%, respectively, similar to that of the genus *Platydema*. All the above genera showed atypically high

G+C content compared to the usually occurring G+C content of 30–50% in insects.

## Discussion

### Efficiency of DNA barcodes in identification of Tenebrionidae species

The results suggest that DNA barcodes are effective in identification of the Tenebrionidae species. For example, a large barcode gap (interspecific variation=20.8%, range 20.7–20.9%, SE=2.1%) was observed between *Tribolium castaneum* and *Tribolium confusum* despite morphological similarities between the two species. Taxonomical identification of the genus *Gonocephalum* belonging to the tribe Opatrini is difficult due to morphological similarities between the species (Kim & Kim 2000). Four species, *G. persimile*, *G. pubens*, *G. coenosum*, and *G. coriaceum*, were sequenced for the COI gene, and the mean interspecific divergences between *G. persimile* and *G. pubens* and between *G. persimile* and *G. coenosum* were 15.4%, whereas those between *G. pubens* and *G. coriaceum* and between *G. coenosum* and *G. coriaceum* were 12.2%. On the other hand, those between *G. pubens* and *G. coenosum* was 0.1%. This result suggests that the classification of the Korean species of *Gonocephalum*, in particular, those of *G. pubens* and *G. coenosum* needs to be reviewed.

Kim (2003) discussed that the genus *Gonocephalum* belonging to the tribe Opatrini is a monophyletic group. However, the genus *Idisia* (*Idisia ornata*) belonging to the same tribe shows a paraphyletic cluster with another genus *Tenebrio*. The DNA barcodes generated in this study also indicate that *Idisia ornata* has a closer genetic similarity with *Tenebrio molitor* than with the genus *Gonocephalum* (data not shown). Similarly, while it is known that both *Phaleromela subhumeralis* in the tribe Diaperinae and the tribe Opatrini inhabit sand dunes, it is also known that their respective subfamilies Diaperinae and Tenebrioninae are genetically closely related (Watt 1974, Doyen 1982).

### Relationship between tenebrionid species and habitat preference

The NJ tree according to habitat preference of the Korean Tenebrionidae species is given in Figure 2. Tenebrionidae species inhabit diverse habitats and the species have adapted to their unique habitat environment. After grouping the species based on habitat preference, the genetic differences and the NJ tree were analyzed (Table 3, Fig. 2).

The results indicated that members of the Korean Tenebrionidae formed a coherent cluster by taxa and habitat preference such as cereals, rotten wood, and macrofungi. The tree shows that the genus *Tribolium* is not monophyletic. The

genus *Gonocephalum* in sand dune habitat formed a monophyletic cluster. The mean of pairwise divergences between interspecies within a group was 22.6% (range 20.7–24.1%, SE=2.2%) for cereal group, 29.0% (range 29.0–29.0%, SE=2.7%) for rotten wood group, 21.6% (range 15.2–26.8%, SE=2.2%) for fungi group, and 8.6% (range 0.0–15.6%, SE=1.1%) for sand dune group. As a conclusion, DNA barcode data obtained from the mitochondrial COI gene could effectively discriminate the species and groups of the Korean Tenebrionidae, which is based on habitat preference, despite of some exceptional cases.

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